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**Introduction**

Arrayit offers the TrayMix™ S4 Microarray Hybridization Station to reduce hybridization times as much as 90% using innovative micro-mixing chaotic advection technology. Excellent experimental results are ensured by completely homogeneous dispersion of the probe molecules across the 21 x 60 mm reaction area. TrayMix™ S4 significantly reduces hybridization time while offering reproducible and robust results from one experiment to the next, using as little as 5 pmole of labeled material in the hybridization mixture. The system is easy to use and maintain, and requires no expensive gaskets or special cover slips. Greater specificity of hybridization is achieved while reducing the coefficients of variation (CV). Enhance the analysis of your results of gene expression, protein expression profiling, mutation detection, comparative genomic hybridization, genotyping, FISH, and many other applications.
**System Features**
Review the current micro fluidic mixing techniques and see the video of the TrayMix™ Mixing loop in action at:
http://arrayit.blogspot.com/2008/10/microarray-hybridization.html

Current micro-fluidic mixing technologies that are being implemented in the microarray industry include, Turbulent Flow, Rotary Mixing with air bubbles, Laminar flow, Acoustic waves, Surface Acoustic Waves, and Chaotic Mixing. The goal of mixing a microarray binding reaction is to assure that every molecule in solution finds its binding partner immobilized on the microarray as quickly as possible. In other words, the most desirable binding reaction has fast and complete diffusion of all biomolecules in solution over all microarray spots and remains a homogeneous mixture until the reaction is complete. The chaotic mixing method of the TrayMix is superior.

Sound microarray data can be achieved using inexpensive hybridization cassettes, however, in certain cases active mixing has been shown to speed up binding reactions, improve data quality, and reduce the number of molecules required in solution for the binding or hybridization reaction. Reasons to use the TrayMix for microarray binding reactions include:
• Save time and money by performing tests faster with less test sample
• Minimize handling of microarrays, which reduces the possibility of human error.
• Get better control over the experimental variables, resulting in increased reproducibility
• Empower users to define, edit and store individual methods and protocols
• Save and link experimental or testing procedures to database

**In a turbulent flow system,** the hybridization cocktail is mixed by the using random contact with the physical structure of a reaction chamber. One challenge of this type of system is obtaining homogeneity of the reaction mixture.

**Rotary mixing** with air bubbles is performed inside a sealed reaction cassette by rotating a trapped air bubble over the microarray. Binding reactions cannot take place in air, only in solution. Therefore a challenge of this approach is minimizing air bubble-based oxidization of the fluorescent dyes commonly used in microarray reactions, which would lead to lower signals and elevated background. Another challenge is that if an air bubble were to become trapped, the reaction in the trapped area would not proceed.

**Laminar Flow** is generated by using a small diaphragm pumping system at each end of the microarray to move the binding reaction sample back and forth across the microarray. Recent micro-fluidic studies show that laminar flow mixing can produce layers of liquid that flow over top of each other, thus one challenge of this approach is to obtain fully homogeneous and complete mixing of sample during the laminar flow process.

**Surface Acoustic Waves** generated by piezoelectric transducers are used to cause streaming of the hybridization reactions under cover slips or lifter slips. Some of the same challenges that apply to laminar flow also apply to systems that use surface acoustic waves.

**Chaotic Advection Mixing** used by the TrayMix is accomplished using micro fluidic pumps and a mixing loop. The overall movement of liquid in properly configured systems is chaotic due to the extremely complex direction and speed of fluids provided by the mixing loop. This type of system provides the most complete mixing of low volumes of liquid in the shortest amount of time.
**System Specifications**

The hybridization temperature is controllable from 20-100°C +/-0.1°C. Upstream of the reaction area, which includes the mixing loop and hybridization chamber, a manifold introduces a programmable 5-way reagent selection system. Pre-hybridization, hybridization buffer, washing solutions and decontamination solution are all computer controlled and programmable. Substrate slide drying is achieved after hybridization and washing using the ArrayIt® High-Speed Centrifuge, which dries the substrate slide completely in less than 10 seconds in preparation for microarray scanning.
## Technical Specifications

<table>
<thead>
<tr>
<th>Specifications</th>
<th>Details</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Dimensions</strong></td>
<td>13.7 x 18.3 x 8.3 in (35 x 26.5 x 14 cm)</td>
</tr>
<tr>
<td><strong>Weight</strong></td>
<td>19 kg</td>
</tr>
<tr>
<td><strong>Capacity</strong></td>
<td>4 standard glass substrate slides, individual or simultaneous</td>
</tr>
<tr>
<td><strong>Microarray substrate</strong></td>
<td>25 x 76 x 1 mm</td>
</tr>
<tr>
<td><strong>Hybridization area</strong></td>
<td>21 x 60 mm</td>
</tr>
</tbody>
</table>

### Hybridization volume

- Hybridization chamber: 60 µl
- Mixing loop: 290 µl

### Hybridization reaction chamber

Specifically designed to maintain consistent reaction volumes from hybridization to hybridization using a shim system. Rubber gaskets are used to seal the chamber, but the gaskets do not define the volume of the hybridization reaction.

### Biological samples

- Optimal quantity: 5 to 50 pmol
- Injected volume via pipette: 30 µl or 60 µL

### Temperature

20-100°C +/-0.1°C
## Buffers and solutions
Chemically-resistant to all standard biochemical reagents

## Programming
PC + BioTray software in a windows operating system. Four stations can be run using a single computer.

## Flexibility
Automation through computer control and easy to use software. The injection port is directly connected to the chamber allowing specific procedures to be implemented including multiple hybridizations, sandwich assays, and enzymatic reactions.

## Repeatability
The computer controlled system performs repeatable tasks much more precisely than manual operations.

## Complete automation
Homogeneous mixing in a controlled reaction. Programmed cycles perform pre-hybridization, hybridization, up to three different wash steps, and complete cleaning of the system in preparation for the subsequent hybridization.

## Cost effective
No expensive cover slips and very easy to maintain. Affordable supporting microarray products from ArrayIt®, the leading brand name in the industry for more than a decade.

### Each hybridization chamber is totally independent. They can be launched simultaneously or programmed to run different protocols using an easy to use graphical user interface.

### Once the system is programmed and the microarray substrate slides are in place on the platform, the system automatically delivers solutions into the hybridization chambers. The 50 µl hybridization chambers are sealed using gaskets located on the upper portion of the lids that each microarray when they are in the closed and locked position.

Up to 60 µl of sample is manually introduced into each hybridization chamber via a dedicated access port. During hybridization, the ports are closed to prevent evaporation.
Software Features and Programming
System programming is achieved via the easy to use graphical user interface shown below.

Set Pre-hybridization Time and Temperature.
Set Hybridization Time and Temperature.
Set Wash Parameters, program is ready to run!
Insert microarray slide, click proceed.
Check the system is locked properly and port cap is properly closed.
Programmed Pre-hybridization Program Starts, is useful to pre-hybridization to get the microarray at proper temperature and to wet the microarray prior to adding the labeled sample.
After Pre-hybridization is completed, temperature is stabilized, the next step is to remove air bubbles for the chaotic advection mixing loop.
Air bubbles are automatically removed.
Injecting the sample is done with a volume of 30 µl or 60 µl, set volume, inject sample and click proceed.
Depending on the volume of injection, the mixing loop automatically compensates for the volume injected for hybridization.
Hybridization proceeds at set time and temperature, with constant mixing.
Wash steps proceed automatically based on set program parameters.
When chamber is empty, microarray is removed from the system and dried in a microarray centrifuge.

The software interface provides several functions:

- Traceability of operations and generation of reports in HTML format
- Saving and loading of protocols
- Flexibility to modify the operation of the system in all of the experimental steps including pre-hybridization, hybridization and washing.
Performance Data

Table 1. Average fluorescence values and coefficients of variation (CVs) measured under different hybridization conditions (2 hours of hybridization).

<table>
<thead>
<tr>
<th>Probe concentration (0.1 µM)</th>
<th>Solution volume (µl)</th>
<th>Probe quantity (pmoles)</th>
<th>Method</th>
<th>Hybridization solution</th>
<th>Fluorescent Mean (a.u.)</th>
<th>CV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Static Hybridization</td>
<td>50</td>
<td>5</td>
<td>cover slip</td>
<td>homogenous</td>
<td>5500</td>
<td>0.33</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td>50</td>
<td>TrayMix™ S2</td>
<td>homogenous</td>
<td>38500</td>
<td>0.56</td>
</tr>
<tr>
<td>Dynamic Hybridization</td>
<td>500</td>
<td>50</td>
<td>TrayMix™ S2</td>
<td>homogenous</td>
<td>11474</td>
<td>0.18</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td>50</td>
<td>TrayMix™ S2</td>
<td>non homogeneous</td>
<td>11823</td>
<td>0.17</td>
</tr>
</tbody>
</table>

Table 2. Average value for fluorescence and CV obtained with the same quantity of targets, both with and without agitation (2 hours of hybridization).

<table>
<thead>
<tr>
<th>Probe quantity (5 pmole)</th>
<th>Solution volume (µL)</th>
<th>Concentration (µM)</th>
<th>Method</th>
<th>Hybridization solution</th>
<th>Fluorescence mean (a.u.)</th>
<th>CV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Static hybridization</td>
<td>50</td>
<td>0.1</td>
<td>cover slip</td>
<td>Homogenous</td>
<td>5500</td>
<td>0.33</td>
</tr>
<tr>
<td>Dynamic hybridization</td>
<td>500</td>
<td>0.01</td>
<td>TrayMix™ S2</td>
<td>Non-homogeneous</td>
<td>6452</td>
<td>0.16</td>
</tr>
</tbody>
</table>

NB: The CV of the system after 2 hours is equivalent to that of static systems with treatment under coverslip for 12 hours.

Signal/Noise vs Specificity
Microarrays hybridized for two hours with a single-stranded CY3 labeled DNA (84 mers) complementary to allele a. Hybridizations were performed with 5 pmol of target under the coverslip method and with the TrayMix™. Allele b was used as a control for hybridization specificity. All spot features were analyzed and compared to the local background signal for each of the experiments. Results comparing the mean signal to noise ratio of the fluorescent CY3 signals to the local background for allele a (black) and allele b (grey) of three experiments ± SD (Standard Deviation).

The figure shows that the specific signal/noise ratio (allele a – black bars) is enhanced from 3.5 to 4.2, while the non-specific signal/noise ratio (allele b - grey bars) is slightly reduced under the dynamic hybridization conditions of the TrayMix™. Comparisons of results after 12 hours of static hybridization are even more favorable to TrayMix™ (Data not shown). Chaotic mixing enhances the hybridization results in all important areas including time, homogeneity, and signal to noise ratio. In addition, the automation of this step is crucial for reproducibility. The system has thus been developed in view of attaining the levels of reproducibility essential for diagnostic purposes. Chaotic advection offers an even more significant advantage to passive hybridization when using high molecular weight molecules in applications such as Microarray Comparative Genomic Hybridization (aCGH).

Conclusion and perspectives
TrayMix™ offers a true innovation for enhancing hybridization efficacy, sensitivity, reproducibility and robustness while easy to use and flexible. The injection port is designed to allow small amounts of new reagents to be automatically and sequentially administered for pre- or post-hybridization processing (e.g. enzymatic reactions, chemical reactions,
sandwich reactions, etc...). This permits conception of more complicated processes such as oligo elongation and multi-step reactions. Furthermore, the technology allows adaptation to either multiple slides or multiple reaction areas per slide. It is to be noted that the TrayMix technology is applicable to all types of microarrays such as, CHIP on Chip, protein and peptide, CGH, and FISH. This vast spectrum of applications makes the TrayMix™ a great investment of anyone processing microarrays.

Technical Support
Please direct technical questions to arrayit@arrayit.com or call 408-744-1331.
## Ordering Information

<table>
<thead>
<tr>
<th>Product</th>
<th>Description</th>
<th>Catalog ID</th>
<th>Price (US dollars)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>TrayMix™ S4 Hybridization Station</td>
<td>Microarray hybridization station for 4 microarrays per cycle based on chaotic advection mixing technology. Each system includes a one year warranty, one year of complimentary software upgrades and complete technical support.</td>
<td>TMHS</td>
<td>$36,535</td>
</tr>
<tr>
<td>ArrayIt Microarray Hybridization, Processing and Hardware System</td>
<td>ArrayIt complete system for DNA and protein microarray hybridization and processing, circulating water bath with digital temperature control, magnetic stir plates with heating (2 each), single substrate slide submersible hybridization cassettes (5 each), high-throughput wash stations (5 each), microarray high-speed centrifuge, microarray air jet with oil-free compressor and 0.1 µm air filter, digital platform mixer with 0-1,000 rpm control and 0-99 min timer, microarray inspection microscope with 30X objective, binocular eyepieces and LED illumination, microarray pin and printhead cleaning kit with ultrasonic bath, pin holder and cleaning reagents, and microarray microplate centrifuge with 5,000 rpm and 1,500 x g centrifugal force.</td>
<td>APHHS</td>
<td>$16,495</td>
</tr>
<tr>
<td>Ozone Free Box</td>
<td>ArrayIt® InnoScan® Ozone Free Box™ reduces ozone levels to 1 part-per-billion to eliminate ozone-mediated degradation of Arrayit Green540 and Red640, cyanine 3 and cyanine 5, and other dyes used for microarray hybridization. For use with the TrayMix™ S4 Hybridization Station.</td>
<td>OFB100</td>
<td>$18,900</td>
</tr>
</tbody>
</table>

*Please contact us directly for exact pricing on these products by email arrayit@arrayit.com, telephone (408) 744-1331, Fax (408) 744-1711, or click purchase button above to proceed directly to the purchase page.

*International pricing may vary as much as 30% (or more depending on country) due to import duties, stocking fees and technical support.

*Add shipping and handling to all orders.
Warranty
ArrayIt® brand products are sold for research purposes. Extreme care and exact attention should be practiced in the use of the materials described herein. All Arrayit products are subject to extensive quality control and are guaranteed to perform as described when used properly. Any problems with these products should be reported to Arrayit immediately. Our liability is limited to the replacement of the product, or a full refund. Any misuse of this product is the full responsibility of the user, and Arrayit makes no warranty or guarantee under such circumstances.